



# EVALUATED WHITE CORN *SORGHUM BICOLOR* TREATMENT BY ENZYME LIGNIN PEROXIDASE INSTEAD OF YELLOW CORN ON $\alpha$ -AMYLASE ACTIVITY IN *CYPRINUS CARPIO* L.

Ali H. H. Al-Gharrawi<sup>1</sup>, Dhamyaa O. A. A-Saadi<sup>2</sup> and Mohammad H. Alasha' ab<sup>3</sup>

<sup>1</sup>Al-Mammon College, Iraq.

<sup>2</sup>College of Agricultural Engineering Sciences, University of Baghdad, Iraq.

<sup>3</sup>Ministry of Science and Technology, Agricultural Researches Directorate, Iraq.

## Abstract

The study was conducted to investigate the effect of replacing the white corn after improving their nutritional value by reducing the tannins by Lignin peroxidase enzyme which locality isolated from bacteria *Spreotpmyce* ssp. and substituted for yellow corn in *Cyprinus carpio* L. diets on  $\alpha$ -amylase activity. The fish average weight ( $25.12 \pm 3.75$  gm/fish) fed on seven experimental diets contain two substitution levels of white corn for yellow corn, T<sub>1</sub> and T<sub>2</sub> used white corn without treatment substitution 50% and 100% for yellow corn, T<sub>3</sub> and T<sub>4</sub> used white corn treatment before grinded by enzyme at same substitution level, T<sub>5</sub> and T<sub>6</sub> used white corn treatment after grinded by enzyme at same substitution and T<sub>7</sub> without white corn for control. The intestinal fish distributed into 3 parts (foreground, middle part, end part). The results of the experiment showed the distribution of the  $\alpha$ -amylase enzyme in all the gastrointestinal tract (foreground, middle part, end part), the highest effectiveness was in the foreground and middle part of the intestine which significant different  $p \leq 0.05$  with end part, it were before starvation (1.89, 1.77 and 0.55 U/mg protein at the nutrition), and the vale after starvation 5 day 0.546, 0.554 and 0.04 U/mg protein at the nutrition respectively. The results also showed the significant value of enzyme activity in the foreground and middle part compared to end parts after 30 days fish feeding. While there were a significant difference of amylase activity between T<sub>1</sub> and T<sub>2</sub> compared with all other treatments. The value recorded (1.146, 1.156, 1.397, 1.501, 1.511, 1.498 and 1.498 U/ mg protein at the nutrition) for foreground and (1.158, 1.149, 1.445, 1.499, 1.498, 1.487 and 1.476 U/ mg protein at the nutrition) for middle part for all treatment respectively. These data suggest, it is possible to replace white corn treatment with lignin peroxidase enzyme up to 100% instead of yellow corn a without any effect on amylase activity.

**Key word** : enzyme, amylase, digestive, carp, white corn.

## Introduction

The Amylase enzyme belongs to a group of hydrolase enzymes. The amylase enzyme compose of a number of enzymes similar to a-amylase (b-amylase, gluco-amylase, etc.) all of which are involved in the analysis of the glycosidic bond in polysaccharides. However, they differ in the mechanism of action (Kujawski *et al.*, 2002). The a-amylase enzyme is highly specialized and it does not analyze cellulose, gums excepted the sugar have glucose which associated with each other with the glycosidic bonds a-1, 4 (Pandy *et al.*, 2000). Animal a-amylase can be obtained from saliva, urine and blood and animal pancreas is the primary source of animal amylase. Kerr (1950). As for omnivorous fish, the enzyme is concentrated

in the pancreas and it is found that the amounts of this enzyme vary depending on the type of fish and the way they are fed (Hidalgo *et al.*, 1999). The (Rungkan *et al.*, 2009) work to isolated the a-amylase from the gut of nile tilapia *Oreochromis niloticus*. The researchers said in the field of fish nutrition that the common carp fish *Cyprinus carpio* L. characterized by containing high amounts of a-amylase (Kuzmina *et al.*, 2003). The researcher (Chakrabarti *et al.*, 1995) noted the enzymes were spread throughout the digestive system and found in most of its parts when studying eleven species of fish. They stressed (Kapoor *et al.*, 1976) that the levels of a-amylase enzyme in the digestive system of the common carp fish is much higher than levels in other fish, which can benefit from the carbohydrates. In contrast to the



**Table 2:** Chemical composition for experimental diets (Calculated by dry matter %).

T <sub>7</sub>	T <sub>6</sub>	T <sub>5</sub>	T <sub>4</sub>	T <sub>3</sub>	T <sub>2</sub>	T <sub>1</sub>	Nutrition constituents for diets
28.14	28.51	28.22	28.74	28.34	29.04	28.38	Crud Protein
6.73	6.57	6.88	6.29	6.66	6.75	6.81	Ether Extract
7.66	6.19	6.47	6.65	6.35	6.22	6.12	Ash
7.41	7.49	7.58	7.32	7.19	7.84	7.21	Crud fiber
50.06	51.24	50.85	51.00	51.46	50.51	51.48	*NFE
1445.61	1463.19	1462.75	1454.83	1469.12	1466.05	1472.11	** Metabolic EnergyKJ/Kg

hours and at temperature of -18 m. The fatty layer was removed the parts mixed with the extraction solution of potassium phosphate solution at ratio (1:4, weight: volume), homogeneous by electric mixture for 2 minute, the mixture transfer to glass container and mix by using a magnetic stirrer at 4<sup>o</sup>C for 6 hours. The mixture was mediated by a piece of filler to remove the excess parts. The cold centrifugal process was performed at a speed of 10000 rpm for 30 minutes to dispose of the remaining parts, the supernatural was separated from the precipitate and the stink is obtained and the enzymatic efficiency estimated by the spectrophotometer at wave length of 540 nano meters. AL-Quraishi (2005).

**Statistical Analysis**

The statistical analysis was carried by using the steps of the general statistical model (SAS) Statistical Analysis System (SAS). Sas (2012) and determined the effect of the coefficients of the studied properties using the complete randomized design (CRD). The averages were compared to each characteristic using a Duncan test at a probability level of P≤0.05 to determine mean differences between average transactions. Duncan (1955).

**Results and Discussion**

The results of the study to estimate the enzymatic efficacy of α-amylase in the crude extract of the gastrointestinal tract of *C. carpio* showed that the enzyme was spread along the gastrointestinal tract after starvation and feeding of fish. The studied parts recorded enzymatic efficiency at starvation for 5 days and were

0.546, 0.554 and 0.04 mg/mg protein, while those values in fish fed on the diet increased to 1.89, 1.77 and 0.55 mg/mg for the studied front, middle and back parts respectively. The results of the statistical analysis showed no significant differences between the front and middle parts, which differed significantly P≤0.05 with the back part (Table 3).

Table 4, showed the enzymatic efficiency of the amylase enzyme for the gastrointestinal tract extract after 30 days of feeding the fish on diets containing different percentages of white corn

with different treatments. The values were significantly higher than the values recorded in starvation, which differed significantly P≤0.05 (Table 3). The results of statistical analysis showed significant differences p≤0.05 between T<sub>1</sub> and T<sub>2</sub> for all treatments for the foreground and middle sections of the enzyme activity were 1.498 and 1.498 units/mg of protein. The values of 1.146, 1.156, 1.397, 11.50, 1.511, 1.498 and 1.498 (P≤0.05) with the end for all the treatments (Table 4).

The difficulty of studying the efficiency of digestive enzymes in fish was strongly related to the method of collection and efficiency evaluation. Therefore, the results differed according to these methods. Some researchers used the entire digestive tract, others divided it into parts, others the lining of the intestines and the difference of the substrate AL-Aldibikl (1996). In general, α-amylase activity increases in diets with high carbohydrate levels in general and starch in particular (Fountoulaki *et al.*, 2005). The results of the current study showed the prevalence of α-amylase along the gastrointestinal tract in all studied parts (foreground, middle and end), the highest in the foreground and middle part compared to the end part (Table 3). These results were consistent with the results of (Frais *et al.*, 1981) studies in the study of the distribution of digestive enzymes of catfish *Ameiurus nebulosus* and common carp *C. carpio* L. that the enzyme was spread along the gastrointestinal tract and that the activity of the enzyme increases with the increase of intestinal filling of food, the enzyme's effectiveness in feeding evaluated it in starvation. The

**Table 3:** The level of effective of α-amylase enzyme before and after starvation.

Significantly level	(Mean±standard deviation)		Part of gastrointestinal tract
	After Starvation for 5 Dsys	Before Starvation	
*	0.546±0.04 bA	1.89±0.08 Aa	Foreground part
*	0.554±0.04 bA	1.77±0.06 aA	Middle part
*	0.04±0.00 aB	0.55±0.02 Ba	Back part

amylase enzyme was associated with the chyme produces by mechanical and chemical effects of the leaf, consisting of partially digested food, water and various digestive enzymes, the chyme pass slowly through the pyloric sphincter, the intestine will digest food to the chyme in any where ranges from 40 minutes to 3 hours at most and is associated with the lining of the intestinal wall of

**Table 4:** Effect of the level and treatment of white corn and the section (gastrointestinal tract) The efficiency level of  $\alpha$ -amylase enzyme.

Significantly level	Control without White corn	The efficiency, Unite/mg protein (after 30 days from the start nutrition)						The part of gastrointestinal tract
		White corn treatment by enzyme after milling		White corn treatment by enzyme before milling		White corn without treatment		
		T <sub>7</sub>	T <sub>6</sub>	T <sub>5</sub>	T <sub>4</sub>	T <sub>3</sub>	T <sub>2</sub>	
*	1.498±0.07aA	1.498±0.05aA	1.511±0.02aA	1.501±0.02aA	1.397±0.05aA	1.156±0.03bA	1.146±0.05Ab	Foreground part
*	1.476±0.05Aa	1.487±0.03Aa	1.498±0.02Aa	1.499±0.02Aa	1.445±0.05aA	1.149±0.05Ab	1.158±0.07bA	Middle part
*	0.432±0.03Bb	0.522±0.06Bb	0.501±0.04Bb	0.533±0.04Bb	0.543±0.03Bb	0.345±0.04Bb	0.364±0.05Bb	End part

the mucus membrane (Hoehne *et al.*, 2001). This explains the distribution of the enzyme along the gastrointestinal tract in this study. The results of this study were similar to the results of the study (Miriam *et al.*, 2002) when starving and re feeding of the sturgeon *Acipenser naccric* and trout *Oncorhynchus mykiss* that the effectiveness of the enzyme was spread along the digestive tract and the highest value was in the foreground and middle of the digestive tract. The results of the current study (Chakarabarti *et al.*, 2006) confirmed that the enzyme amylase is spread along the gastrointestinal tract of the silver carp *Hypophthalmichthys molitrix* and higher in the front and middle part. In his study, the researcher noted the prevalence of the digestive enzymes of *C. carpio* L. along the gastrointestinal tract at higher rates in the foreground and middle part of the gastrointestinal tract compared with the end part.

The effect of replacing white corn with yellow corn in this study when feeding common carp *C. carpio* L. on diets containing different percentages of white corn, there was a significant difference  $P \leq 0.05$  in the enzyme efficiency values of the first and second treatments with all other treatments and the control treatment (Table 4). This may be due to equal the carbohydrate ratios (Table 2), that support the  $\alpha$ -amylase activity depend on carbohydrate levels in general and starch in particular. The results of this study were confirmed by (Salah *et al.*, 2016) when they studied the effectiveness of  $\alpha$ -amylase for three species of fish, *Lucius vorax* (carnivores) and *Liza abu* (omnivores) and blue tilapia *Oreochromis aureus* (herbivores) in southern of Iraq (Hor al-Jabbaysh) that the efficiency of  $\alpha$ -amylase was clear in tilapia about other fish because it was a herbivores. In their study of the functions of digestive enzymes and their relationship with food in the silver carp *Hypophthalmichthys molitrix* and big head carp *Hypophthalmichthys nobilis*, the enzyme was the highest in fish fed on high carbohydrate diets. The (Baohua *et al.*, 2009) explained when feeding common carp *C. carpio* L. on three diets containing 35% protein supplemented with 1.5, 2.1 and 3.2 mg/kg of xylolgo saccharides (XOS) for three treatments, the

efficiency of amylase was significantly higher when addition 3.2 mg/kg of XOS compared to the control treatment without supplementation XOS. The efficiency of  $\alpha$ -amylase did not have any significant effect when substituting soybeans with grass pea seed *Lathyrus sativus* L. which heat-treated in common carp diets and may be due to the approximate carbohydrate ratios in the used diets (AL-Gharrawi *et al.*, 2016).

## Reference

- Kujawski, M., R. Ziobro and H. Gambus (2002). Raw starch degradation by pullulanase. *Acta Technology Alimentaria.*, **1(2)**: 3-35.
- Pandy, A., P. Nigam, C.R. Soccal, V.T. Soccal, D. Sing and R. Mohan (2000). Advances in microbial amylase. *Biotechnol. Appl. Biochem.*, **31**: 135-152.
- Kerr, R.W. (1950). Chemistry and Industry of starch. AP, Inc. New York.
- Hidalgo, M.C., E. Urea and A. Sanz (1999). Comparative study of digestive enzyme in fish with different nutritional habits photolytic and amylase activities. *Aquaculture.*, **170(3-4)**: 267-283.
- Rungkan, K., A. Nontawith and E. Arunee (2009). Characterization and activity of digestive enzymes in different size of Nile Tilapia. *Kasetsart. J. Nat. Sci.*, **43**: 143-153.
- Kuzmina, V., L. Glatman and A. Gelman (2003). Amlolytic activity in fish intestinal mucosa: Temperature eff. Abstract. *Comp. Biochem. Physiol. Biochem. Mol. Biol.*, **134(3)**: 529-534.
- Chakarabarti, I., A. MD. Gan, K.K. Chaki, R. Sur and K.K. Misra (1995). Digestive enzyme in 11 freshwater teleost fish species in relation to food habit and niche segregation. *Comp. Biochem. Physiol. Vol.*, **112A(1)**: 167-177.
- Kapoor, B.G., H. Smith and I.A. Verighina (1976). The alimentary canal and digestive in telests. *Adv. Mar. Biol.* **13**:109-239.
- Krogdahi, A. and A.M. Bakke-Mckellep (2005). Fasting and refeeding cause rapid changes in intestinal tissue mass and digestive enzyme capacities of Atlantic Salmon *Salmo salar* L. *Comp. Biochem. Physiol.*, **141**: 450-460.
- Al-Gharrawi, Ali Hussein Hassan (2012). The relationship of the type of diet to the enzymes of common carp *Cyprinus carpio* L. and its impact on growth and living. P.hD. thesis, College of Agriculture, University of Baghdad, 125.

- Smith, R.R. (1971). A method for measuring digestibility metabolizable energy of feed. *Prog. Fish. Cult.*, **33**:132-134.
- AOAC (1990). Official Methods of Analysis, 15<sup>th</sup> edition, Association of Official Analytical Chemists, Washington DC.
- Al-Quraishi, Abdel-Al Farhan (2005). Extraction of  $\alpha$ -amylase enzyme from the hepatic pancreas of common carp fish, purification and study of some of its properties and applied applications, Master Thesis, College of Agriculture, University of Baghdad.
- SAS (2012). Statistical Analysis System, User's Guide. Statistical. Version 9. I<sup>st</sup> ed. SAS. Inst. Inc. Cary. N.C. USA.
- Duncan, D.B. (1955). Multiple Rang and Multiple F-test. *Biometrics.*, **11**: 4-42
- Al-Tamimi, R. Adnan (1998). Effect of protein-to-energy ratio in diets on growth of common carp fingerlings. Master Thesis-Faculty of Agriculture- Basrah University. 64.
- Al-Aldibikl, A. Yacob (1996). Nutritional and metabolic study of small brown fish *Barbus sharpy* and Alcantan *B. xanthopterus* and common carp *Cyprins carpio* L. under different environmental conditions. P.hD. College of Agriculture, University of Basra. 119.
- Fountoulaki, E., M.N. Alexis I. Nengas and B. Venou (2005). Effect of diet composition on nutrient digestibility and digestive enzyme levels of gilthead sea bream *Sparus aurata*. *Aquacult. Res.* **36**: 1-9.
- Frais, M., N.Y. Woo, J. Noiaillac-Depeyre and J.C. Murat (1981). Distribution pattern of digestive enzyme activities in the catfish *Ameiurus nebulosus* and the common carp *Cyprinus carpio* L. *Comp. boil. Physiol.*, **70**: 433-446.
- Hoehne, R.K., E. Kjorsvik and D.R. Gjellesvi (2001). Development of bile salt-dependent lipase in larva turbot. *J. Fish Biol.*, **58**: 737-745.
- Miriam, F., G.G. Manuel, M.H. Cramen, E.M. Amalia, D. Alberto, D. Julio and S. Ana (2002). Effect of starvation and refeeding on digestive enzyme activity in trout *Oncohynchus mykiss* *Comparative Biochemistry and Physiol.*, **149**: 402-425.
- Chakarabarit, R.M., P. Rathore, Mittal and S. Kumar (2006). Functional change in digestive enzyme and characterization of protease of sliver carp and bighead carp hybrid, during early development. *Aquaculture.*, **253**: 694-702 .
- Saleh, O. Abdul Hadi, Aldibikl, A. Yacoub Abdul and J. Mohsen (2016). Estimation of  $\alpha$ -amylase activity in the digestive tract extract of three species of fish in the southern part of Horal-Chabaish. *Basrah Journal of Agricultural Sciences.*, **29(1)**: 286- 295.
- Baohua, X.U., W. Yanbo and L.I. Jianrong (2009). Effect of prebiotic Xylooligosaccharides on growth performances and digestive enzyme activities of allogynogenetic crucin golden carp *Carassius auratus gibelio*. *Fish Physiol. Biochem.*, **35**: 351-357.
- Al-Gharrawi, A.H. Hassan, Al-Ashaab, M. Habas, Jalil and M. Jawad (2016). Effect of various thermal treatments of the seeds of the *Lathyrus sativa* on the secretion of some digestive enzymes of common carp fish. *Cprinus carpio* L.